Transgenic Mouse Facility

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LEADERSHIP & MISSION





Grant MacGregor, DPhil S Director

Shimako Kawauchi, PhD Manager

TMF facilitates use of the mouse as a mammalian experimental system to investigate mechanisms of oncogenesis and testing of cancer therapeutics

- Advises investigators wishing to use genetically engineered mouse models (GEMMs) in their research program, on experimental design and analysis, helps write grant proposals and manuscripts and provides letters of support
- Provides access to specialized expertise and equipment to develop GEMMs, provides technical support, and sources additional reagents required to manipulate the mouse genome and analyze the consequences thereof
- Communicates awareness of novel mouse-related resources, facilitates their acquisition, and provides practical assistance with their use
- Assists researchers by importing, or helping to develop, new experimental approaches necessary to address specific experimental questions in their research

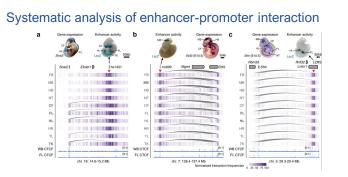
SERVICES, TECHNOLOGIES & EQUIPMENT

Services cover design, development, re-derivation, cryopreservation, and reanimation of GEMMs in an efficient and cost-effective manner, including:

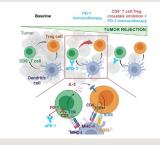
- Consultation, at <u>no cost to PI</u>, on strategies to engineer the mouse genome
- Design and targeted engineering of loci in mouse zygotes via CRISPR (>300 projects completed to date)
- Targeted transgenesis at the Hipp11 and ROSA26 loci
- Targeted engineering of endogenous loci in mES cells including CRISPRmediated humanized gene replacement
- Southern analysis of targeted loci in ES cells and animals including PFGE

- Insertion of conventional multi-copy transgenes and bacterial artificial chromosomes (BAC) at random loci via pronuclear injection of DNA
- Development of RT-PCR genotyping assays
- High-throughput analysis of standard PCR assays using Fragment Analyzer
- Production of large cohorts of genetically defined mice
- Content and figures for grant proposals and manuscripts, letters of support, etc. at <u>no cost to PI</u>

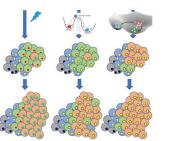
RESEARCH HIGHLIGHTS

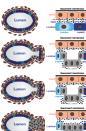


Overcoming limits to immunotherapy in melanoma



NEW MPI P01: Tipping Points in Cancer





\$3.5M Supported CFCCC Members Received 4 New Cancer-relevant Grants (Total Direct Costs) Support Led to 2 New Cancer-Relevant Publications (75%) in IF ≥ 10 Journals BASE SR FUNDING 10% Grant CFCCC Members (75%) BASE SR FUNDING CCSG Recharge Other \$868,145 Annual Budget

TRAINING

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- Personalized training via e-mail and meetings
- Annual lecture on GEMM and genome engineering
- Provision of GEMM and cancer-related resources via TMF website
- TMF workshop idea survey
 Image: Comparison of the survey

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- Recorded tutorials via TMF website

FUTURE PLANS

 In person workshops to provide training in best practices for mouse handling, ID, genotyping, breeding, nomenclature, cryopreservation, genetic background and monitoring, genotyping assays

Internal Advisory Committee

Extensive advocation for CFCCC Membership



Aimee Edinger, VMD, PhD Deputy Associate Director Basic Science, CFCCC



Evgeny Kvon, PhD Assistant Professor Development & Cell Biology



Marcus Seldin, PhD Assistant Professor Biological Chemistry



Claire Lindsell, PhD, BVSc Director, ULAR



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MEMBERS

- The internal advisory committee includes experts in cancer, metabolism systems genetics, mouse biology, mouse genetics and transgenesis
- Member Responsibilities: Review TMF technology, operations, priorities, efficiencies. Provide ideas about GEMMrelated emerging technologies of potential broad use by CFCCC membership
- Selection Process: Identified and petitioned by TMF Scientific Director, using following criteria: CFCCC membership; expertise in use of genetically modified mice in biomedical research; expertise in mouse biology and health, expertise in administrative oversight
- Appointment Terms: Renewed on continuous basis

FREQUENCY Normally annually

FUNCTION

Independent feedback on TMF services and activities

AUTHORITY

Advise on strategic goals, identify opportunities and address challenges

Services, Technologies & Equipment

Design, development, import, re-derivation, cryopreservation, re-animation and production of GEMMs



 Bioinformatic analyses of mouse and human genomics to design strategies for genome engineering



 Microinjection, electroporation and culture of zygotes / preimplantation embryos (two systems)



 Freezing, cryogenic storage and reanimation of sperm, embryos, mES cell lines (multiple freezers with duplicate storage in two buildings)

TagMan, rhAMP based

PCR systems

genotyping via two Bio-Rad RT-

- PFGE and Southern analysis using Bio-Rad CHEF Mapper

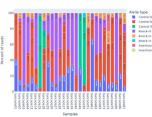




 IVF-based mouse production (multiple incubators) for rapid allele assembly at multiple loci and cohort development of experimental and control animals



Multiple animal holding rooms with ventilated cage racks and sterile caging



- Deeper and faster CRIPSR modification analysis with ONT sequencing and DAJIN2 software
- Tissue culture suite with incubators, hoods and electroporation apparatus (not shown)



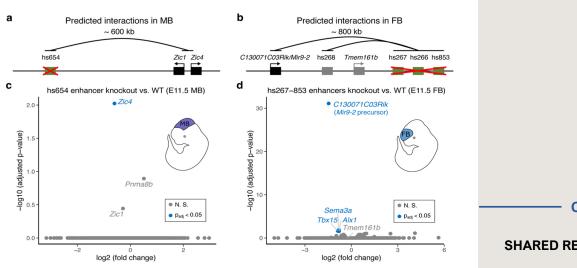
 High-throughput (3 x 96well tray) analysis of standard PCR reactions using Agilent capillary array Fragment Analyzers (two instruments)

CATCHMENT AREA RELEVANCE

Systematic analysis of enhancer-promoter interaction in vivo

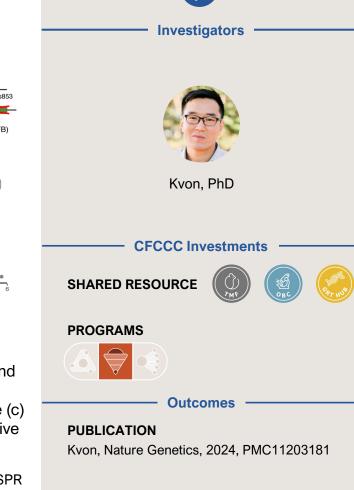
Functional analysis of tissue-specific enhancer-promoter interactions

- Deregulated gene expression can drive oncogenesise.g. loss of expression of tumor suppressors or gain of function of oncogenes
- Dynamics and mechanisms of enhancer-promoter interactions regulating gene expression are poorly understood
- Systematic Hi-C analysis of ~1000 promoter-enhancer interactions in multiple organs during mouse development reveals nearest promoters are frequently bypassed and active enhancers contact each other in clusters



CRISPR-mediated knockout analysis of hs654 and hs267/hs266/hs853 enhancers in mice. a,b, Predicted chromatin interactions between enhancers (green boxes) and target genes (black boxes). c,d, Transcriptome-wide mRNA expression changes in the midbrain of hs654-knockout mice (c) and forebrain of hs267/hs266/hs853-knockout mice (d) relative to wild-type mice

TMF generated numerous lines of KO and KI mice for project via CRISPR



GRANTS DP2 GM149555

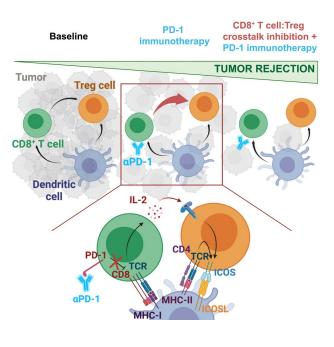
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CATCHMENT AREA RELEVANCE

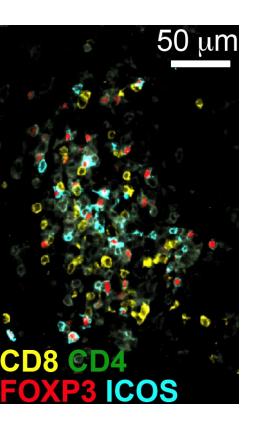
Disrupting CD8 and T signaling enhances immunotherapy

Cutting the phone cable between CD8 and T regulatory cells

- PD-1 blockade expands Tregs in melanoma and limits the efficacy of immunotherapy
- Treg-intrinsic PD-1 inhibition does not cause tumor Treg accumulations
- Anti-PD-1 increases Treg numbers via an intratumor CD8+ T cell/ IL-2/ ICOS axis
- Inhibition of the CD8 + T cell: Treg crosstalk by anti-ICOSL synergizes with anti-PD-1 therapy
- Risk/SDH- This work shows one mechanism of resistance to PD-1 immunotherapy. PD-1 blockade causes the accumulation of tumor Tregs, and restricts its own antitumor efficacy, Inhibition of this mechanism ameliorates PD-1 immunotherapy



TMF imported strains of mice infected with MVM from investigator in Japan, rederived these, and used assisted reproduction to accelerate development of *Pdcd1* ^{loxP}; *Foxp3* ^{CreERT2} and other models for study



Investigators





Marangoni, PhD

Ganesan, MD, PhD





Othy, PhD

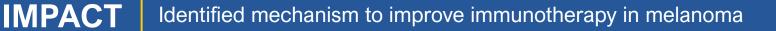
Nie, PhD



PUBLICATION

Marangoni, Cancer Cell, 2024, PMC11285091

GRANTS MRA 929155 DOD ME220176P1



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CATCHMENT AREA RELEVANCE

Investigators

Use of assisted reproduction to accelerate research using GEMMS

Examining Basel-luminal progenitor cell expansion in breast tumors

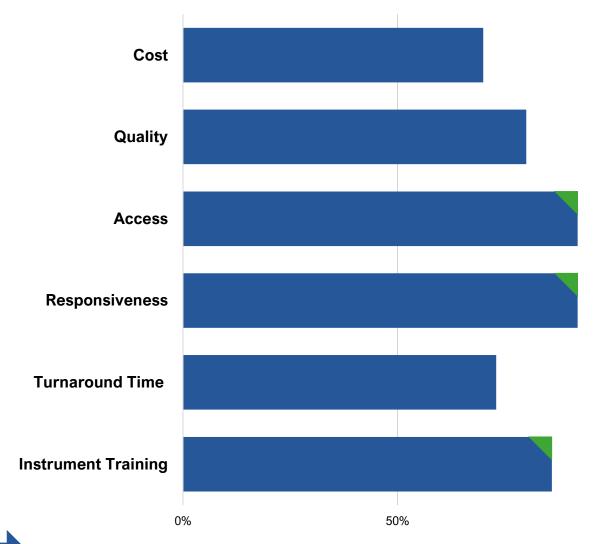
GEMMs - minimum time requried for genotype assembly and cohort production via conventional breeding vs assisted reproduction - e.g. four alleles Brca1^{loxP/loxP}: Trp53^{loxP/loxP}: Fst^{loxP/loxP}: ± Wap-Cre Tg Lawson, PhD Lander, MD, PhD <u>Chr</u> Starting strain genotypes *Brca1*^{loxP/+} (Jax cryorecovery) 11, 70Mb Trp53^{loxP/loxP} (Jax) 11, 101Mb Conventional Assisted Fst^{loxP/loxP} (import) 13, 115Mb Reproduction breeding. Wap-cre +ve (Jax cryorecovery) ? min. "crosses" *min*. crosses Lowengrub, PhD Van Etten, MD, PhD & time & time Genotype assembly **CFCCC** Investments Brca1^{loxP/loxP}. Trp53^{loxP/loxP}; Fst^{loxP/}loxP; Wap-cre + $6 \operatorname{cross} x 3 \operatorname{mo} = 18 \operatorname{mo}$ 6 IVF x 1.5 - 2.5 mo = 9-15 mo Brca1^{loxP/loxP}, Trp53^{loxP/loxP}: Fst^{loxP/loxP} 5 IVF x 1.5 - 2.5 mo = 7.5-12.5 mo $5 \operatorname{cross} x 3 \operatorname{mo} = 15 \operatorname{mo}$ SHARED RESOURCE (B6J first litters often lost) Genotype expansion $3 \operatorname{cross} x 3 \operatorname{mo} = 9 \operatorname{mo}$ 3 IVF x 1.5 - 2.5 mo = 4.5 - 7 moDOT FUNDING PROGRAMS (B6J first litters often lost) 2020, 2022 Cohort Production (c. 50 females ea) 2023 Brca1^{loxP/loxP}, Trp53^{loxP/loxP}; Fst^{loxP/}loxP; Wap-cre + 2 -4 gen x 3 mo = ~9 mo 3 IVF = 2 mo& (B6J first litters often lost) **Outcomes** Brca1^{loxP/loxP}. Trp53^{loxP/loxP}: Fst^{loxP/loxP} PUBLICATION Total time estimate 18 + 9 + 9 = 36 mo 15 + 7 + 2 = 24 moIn Process

Generation of cohorts of GEMMs with complex genotypes via assisted reprodution saves time and animals

GRANTS P01 Tipping Points in Cancer *Pending NOA*

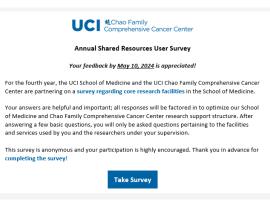
2024 Annual Shared Resource Survey

Excellent + Good (No scores below average received) Therefore 2021



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SURVEY PROMOTION





2024 Core Facilities Survey

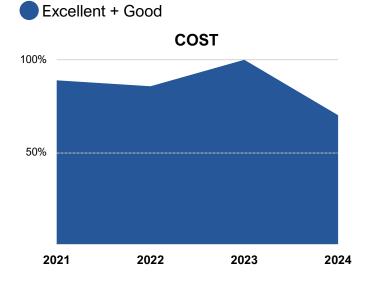
UCI School of Medicine and the UCI Chao Family Comprehensive Cancer Center are partnering on a survey regarding core research facilities in the School of Medicine. Your answers are helpful and important; all responses will be factored in to optimize our research support structure. After answering a few basic questions, you will only be asked questions pertaining to the facilities and services used by you and the researchers under your supervision. This survey Is anonymous. For questions, contact **Claire Brainard Draper**. Please complete the survey **by Mey 10**, 2024.

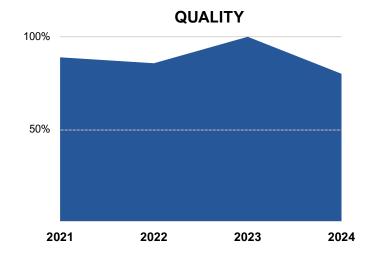


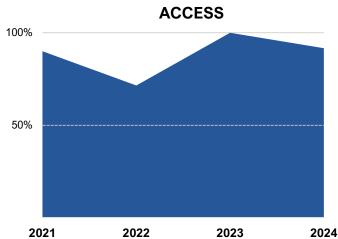
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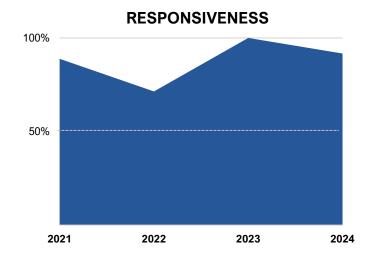
Annual Core Research Facilities Survey

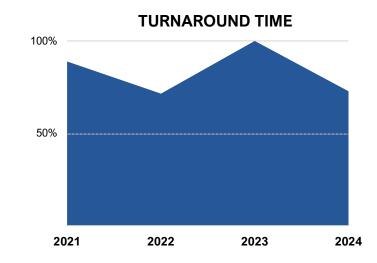


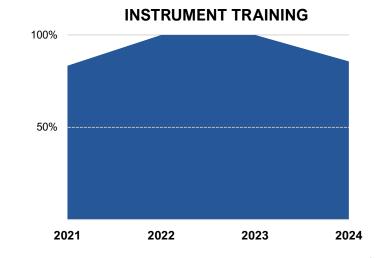












Selected 2024 Publications



| CFCCC INVESTIGATOR(S) | PROGRAM | JOURNAL | YEAR |
|--------------------------|---------|---------------------|---------------|
| Francesco Marangoni, PhD | SPT | | |
| Qing Nie, PhD | SPT | Cancer Cell | 2024 |
| Anand Ganesan, MD, PhD | BIDD | | |
| Evgeny Kvon, PhD | SPT | Nature Genetics | 2024 |
| Qing Nie, PhD | | | |
| Maksim Plikus, PhD | SPT | Nature | 2024 |
| Evgeny Kvon, PhD | | | |
| Evgeny Kvon, PhD | SPT | bioRxiv | 2024 |
| Grant MacGregor, DPhil | SPT | Proc. Natl Acad Sci | in press 2024 |